Group	Strain	Protein(s) Affected	Description	Reference
Parental	14028s	N/A	Wild-type parental strain; highly virulent, highly inflammatory	N/A
Metabolic Attenuation	ΔaroA	3-phosphoshikimate 1- carboxyvinyltransferase	Mutant lacks enzyme connecting glycolysis to aromatic amino acid synthesis; expression of a variety of genes is affected	(1)
Defect in LPS Core	ΔgalE	UDP-galactose-4- epimerase	Mutant lacks the enzyme required to metabolize galactose, a core sugar in LPS	(2, 3)
	ΔrfaG	Glucosyltransferase I	Mutant has highly truncated LPS; no outer core can be added to the inner core; there is no O-antigen	(1, 4)
	ΔrfaH	Transcriptional antiterminator	Mutant has defective core LPS and lacks the O-antigen	(5)
Structural Change to Lipid A	ΔpagP	Palmitoyl transferase for lipid A	Adds a palmitate to lipid A in response to membrane damage; lack of gene results in more hexaacylated LPS (more immunogenic)	(6–9)
	ΔmsbB	(KDO)2-(lauroyl)-lipid IVA acyltransferase	Transfers myristate to lipid A; thought to be a significant factor in attenuation of tumor-targeting VNP20009	(10, 11)
	ΔΙρχΟ	dioxygenase for lipid synthesis	Adds a hydroxyl group to myristate at 3'; altered structure may have implications for TLR4 activation	(9)
	VNP20009	Purl, MsbB	Purl mediates purine biosynthesis; MsbB adds terminal myristate to lipid A. Administered to humans safely in two Phase I clinical trials	(10, 12)
	VNP20009 msbB <sup>+</sup>	(KDO)2-(lauroyl)-lipid IVA acyltransferase	Used to elucidate the role of other mutations in VNP20009	(13)
Defect in O- antigen	Δrfc	O-antigen polymerase	O-antigen is not synthesized to its full length	(5)
	$\Delta rfbK$	phosphomannomutase	Mutant has no O-antigen	(14)
	ΔrfbP	O-antigen transferase	Mutant has no O-antigen	(15)
	∆manA	mannose-6-phosphate isomerase	Mutant is unable to synthesize the O- antigen, but still expresses the full core lipid A	(2)
	ΔrfaL	O-antigen ligase	Mutant has a complete LPS core but lacks the O-antigen	(4)
Other Mutations	ΔmanC	mannose-1-phosphate guanylyltransferase	Mutant produces less LPS, has improved ability to form biofilms, and induces lower reactive oxygen species (ROS) response in macrophages	(16)
	ΔcsgA	cryptic curlin major subunit	CsgA was shown to be a TLR2 PAMP; mutants have reduced production of fimbria	(17, 18)

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